



Identification of some nucleotide mutations in *Waxy* gene (BGIOGA022241) of a mutant rice line

Nguyen Thi Hong^{1,*}, Yoshikazu Tanaka², Vo Thi Minh Tuyen¹, Le Huy Ham¹

¹ Agricultural Genetics Institute, Pham Van Dong, Bac Tu Liem, Hanoi, Vietnam

² The Wakasa-wan Energy Research Center, Fukui, Japan

*Email: nguyenhongdhnn@gmail.com

Abstract: *Waxy* genes of the original variety and its mutant type were sequenced by Sanger method and compared through Nucleotide Basic Local Alignment Search Tool (BLASTN) to clarify differences. BLASTN result showed four nucleotide mutations in coding regions and 59 nucleotide mutations in non-coding regions. Four point mutations in coding regions were: the deletion of T/- at position 34 and the insertion of -T between positions 70 and 71 in exon 3; the substitution of C/T at position 14 in exon 4 and the substitution of T/C at position 115 in exon 9. In 59 mutant nucleotides in non-coding regions, some significant alterations were list: the deletion of nucleotide G at the first of intron 6 and the addition of 32 nucleotides “GGGCCTGCGAAGAAGAACTGGGAGAATGTGCTCCT” at the end of intron 12. For the first trial, a new DNA marker was developed based on the mutation C/T at at position 14 in exon 4 and the substitution of T/C at position 115 in exon 9 to improve efficiency of rice breeding relevant to *Waxy* gene.

Keywords: *Waxy* gene, BGIOGA022241, amylose content, BLASTN, DNA marker development.

I. INTRODUCTION

Rice is a major staple and important food of population in over the world and the demand of it with good quality has been growing in the global market [1]. The endosperm starch is the important feature to assess the quality of rice [2, 3]. Two main properties of endosperm starch are amylose and amylopectine. The amylose is considered the most important predictor of sensory quality of rice and the low content of it is a desirable trait for rice breeding [4]. The synthesis of amylose is controlled mainly by *Waxy* gene, one of key genes [2,5]. Some alleles of *Waxy* gene were listed such as: *Wxa*, *Wxin*, *Wxb*, *Wxop*, and *wx* [6, 7, 8, 9]. Alleles *Wxa* and *Wxop* have been determined in Indica while alleles *Wxin*, *Wxb* and *wx* appeared in Japonica

subspecies [9, 10, 11]. The amylose content of rice endosperm is controlled by complex pathways of the inhibition and the expression of *Waxy* gene through the synthesis of mRNA [12]. It was reported that the accumulation of 3.3 kb *Wx* mRNA (with intron 1) and 2.3 kb *Wx* mRNA (without intron 1) regulates the difference of amylose content [13]. Some mutations published to have effect on the cutting of intron 1 were the insertion of 16 nucleotides [6] or G/T substitution [14] at intron 1/ exon 1 junctions.

The SNP in exon 6, which is identified as *wx* allele, separates varieties with high and intermediate amylose content; and the SNP in exon 4 associating with opaque phenotype, is defined as the *Wx^{op}* allele [9]. Some other SNPs at exon 6 (A/C) and exon 10 (C/T) in

Waxy gene have been determined as the most significant impact on amylose content [15, 16, 17, 18]. Moreover, some changes of *Waxy* gene such as (C/T) SNP at point 2777 in boundary site of intron 7/exon 8 [19]; (A/G) SNP at position 497 from the start codon, leading to the Asp-165/Gly-165 substitution [3]; (G/T) SNP at position 497 (the Arg-158/His-158 substitution in exon 4) and (T/C) SNP at position 595 (Tyr-191/His-191 change in exon5) [20]; (GC/TT) SNP in intron 6, exon 7, intron 7, exon 8 and part of 3' [21] or simple sequence repeats (CT)_(n) and (AATT)_(n) [1]... were considered to effect on amylose content.

It was reported that the diversity of *Waxy* genes among species of *Oryzae* genus was higher than among cultivations in the same subspecies [7]. Spontaneous mutations of *Waxy* gene were found in local rice cultivars from Asian and African countries [4, 10, 22, 23, 24]. In this study, we focused on clarifying the possible changes in *Waxy* gene (BGIOGA022241) between two genotypes belonging to *Oryza Sativa Indica* subspecies: the original variety and its mutant type through some techniques such as: PCR, sequencing and, BLASTN.

II. MATERIALS AND METHOD

A. Materials

Rice samples were the original variety BT62 with high amylose content and its mutant type with lower amylose content.

B. Methods

DNA extraction method: Total DNA of materials were extracted by DNeasy Plant Mini Kit (supplied by QIAGEN) [25].

PCR method: *Waxy* genes of original and mutant varieties were amplified by PCR method. The total 20 µl of PCR reaction

contained 1 µl total DNA (1 ng/µl); 10 µl Prime STAR MAX DNA Polymerase mixture; 0.5 µl forward primer (20 pmol/µl or 20 µM); 0.5 µl reverse primer (20 pmol/µl or 20 µM); 8 µl H₂O. The PCR condition was: 98°C - 2 minutes; 30 cycles of: 98°C - 5 seconds, 60°C - 5 seconds, 72°C - 30 seconds; 72°C - 5 minutes; Keep the sample at 4°C. PCR products were analyzed on a 1.5% agarose gel and purified by QIAquick PCR Purification Kit (supplied by QIAGEN) [25].

Sequencing method: *Waxy* genes of original and mutant varieties were sequenced by Sanger method through the BigDye Terminator Sequencing Standard Kit (Thermofisher) with ABI PRISM 3100 Genetic Analyzer. Sequence reactions were conducted with the cocktail volume of 20 µl including 1 µl DNA (about 20 ng/µl); 4 µl Terminator Ready Reaction Mix; 4 pmol Primer; 11 µl H₂O and the program: 94°C - 2 minutes; 25 cycles of: 96°C -10 seconds, 50°C - 5 seconds, 60°C - 4 minutes; Keep the sample at 4°C. Sequence reactions were purified by DyeEx 2.0 Spin Kit (supplied by QIAGEN) [25] and read by ABI PRISM 3100 Genetic Analyzer.

Analysis method: The sequences of *Waxy* gene of original and mutant varieties were analyzed through Nucleotide Basic Local Alignment Search Tool (BLASTN).

III. RESULTS

A. Primer design and *Waxy* gene (BGIOGA022241) amplification

The sequence of *Waxy* gene (BGIOGA022241) was mined from database of *Oryza sativa Indica* [26]. It locates on chromosome 6 (from 1.931.535 to 1.935.014 forward strand) with 3479 bp including 13 exons and 12 introns (Fig. 1).

IDENTIFICATION OF SOME NUCLEOTIDE MUTATIONS IN WAXY GENE (BGIOSGA022241)...

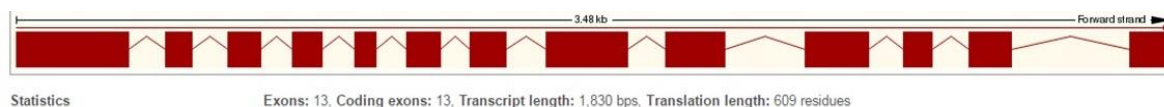


Fig. 1. The structure of *Waxy* gene (BGIOSGA022241) mined from database (Source: <http://www.gramene.org>)

Based on the mined information of *Waxy* gene, eight primer pairs (sixteen primers) were designed to amplify and sequence (Table I).

The full length of the *Waxy* gene (BGIOSGA022241) was amplified by forward primer Wx-1F and reverse primer Wx-8R with the size between 3 kb and 4 kb (Fig. 2).

Table I. The information of primers for *Waxy* gene study.

Name	Sequence (5'-3')	Name	Sequence (5'-3')
Wx-1F	ACAGCAACAGCTAGACAACCACCAT	Wx-5F	AAGTACGACGCAACCACGGTAAGAA
Wx-1R	CTAATCGATCTTGTGATGATCTGA	Wx-5R	GTGGACTAGACGATCTGGGTTCAAA
Wx-2F	TGTGGTGCAATTCATTGCAGATCAA	Wx-6F	TTAGCCGGAAGACCTCTGAGCATTT
Wx-2R	CATCATGGATTCCTTCGAAGAAAGT	Wx-6R	GTAGTGTACCGACTTATCGGTATTA
Wx-3F	TGACAACAGGTGAGGATGTTGTGTT	Wx-7F	GTCTCAGCGTCGACGTAAGCCTATA
Wx-3R	ACGATGGACAGTAGTGCAGGGTTGT	Wx-7R	CCAGTTCTTCGCAGGCCCTGAAAT
Wx-4F	CATCGACGGGTATGAGTAAGATTCT	Wx-8F	GAACAAGACGAACGGTCAAACATGT
Wx-4R	TTCGCCTCGATTGCCTGAAATTTGT	Wx-8R	CATATGTAGATCTCAGGCTCTTCAA

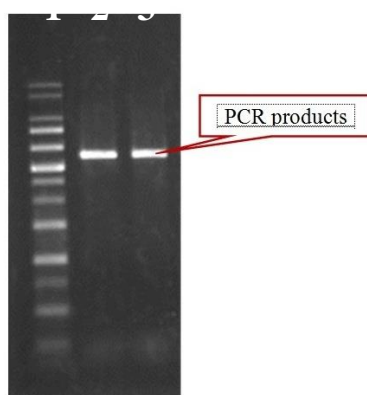


Fig. 2. PCR products of *Waxy* gene on agarose gel 1.5% (1: DNA ladder 1kb; 2: PCR product of the original type; 3: PCR product of mutant type)

B. Sequence the *Waxy* gene (BGIOSGA022241)

The sequencing was conducted by Thermofisher's BigDye Terminator

Sequencing Standard Kit and read by ABI PRISM 3100 Genetic Analyzer and results were shown in Fig. 3.

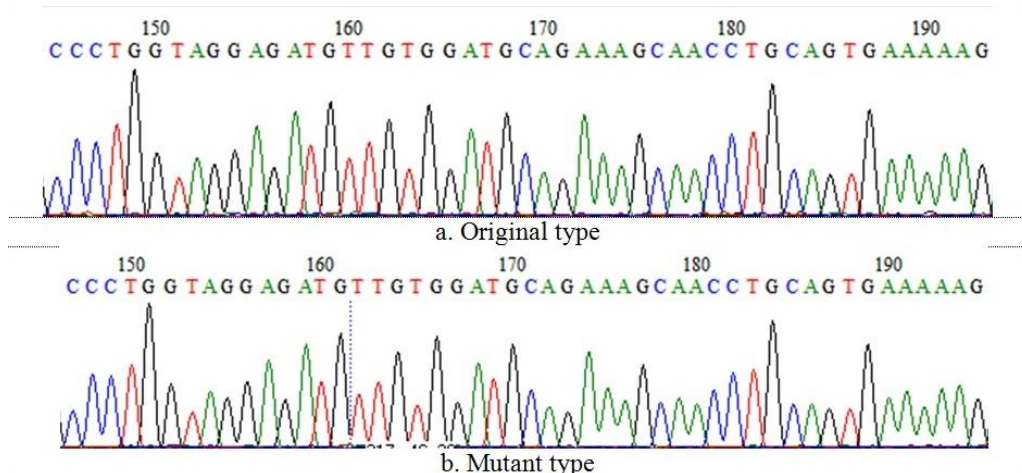


Fig. 3. The result of sequencing *Waxy* gene by Wx-3R primer

(a. Original type – a part sequences of *Waxy* gene in the original type; b. Mutant type - a part sequences of *Waxy* gene in the mutant type)

C. The identification of mutant nucleotides in *Waxy* gene (BGIOGA022241)

The result of comparisons between *Waxy* genes of the original and the mutant variety was shown in Fig. 4; Fig. 5.

```

Query 1 ATCAAGGTTGCAGACAGGTACGAGAGGGTGAGG-TTTTCCATTGCTACAAGCGTGGAGT 59
      |
Sbjct 1 ATCAAGGTTGCAGACAGGTACGAGAGGGTGAGG-TTTTCCATTGCTACAAGCGTGGAGT 60

Query 60 CGACCGTGTGTTTCATCGACCATCCGTCATTCTGGAGAAG 100
      |
Sbjct 61 CGACCGTGTG-TTCATCGACCATCCGTCATTCTGGAGAAG 100
    
```

(a)

```

Query 1 GTTTGGGGAAAGA-TCGGAGAGAAGATCTACGGACCTGACACTGGAGTTGATTACAAAGAC 60
      |
Sbjct 1 GTTTGGGGAAAGACCGGAGAGAAGATCTACGGACCTGACACTGGAGTTGATTACAAAGAC 60

Query 61 AACCAGATGCGTTTCAGCCTTCTTTGCCAG 90
      |
Sbjct 61 AACCAGATGCGTTTCAGCCTTCTTTGCCAG 90
    
```

(b)

```

Query 61 GACAGGAAAATCCCACTGATCGCGTTCATCGGCAGGCTGGAGGAACAGAAGGGCCTGAC 120
      |
Sbjct 61 GACAGGAAAATCCCACTGATCGCGTTCATCGGCAGGCTGGAGGAACAGAAGGGCCTGAC 120

Query 121 GTCATGGCCGCCCATCCCGGAGCTCATGCAGGAGGACGTCAGATCGTTCCTCTG 177
      |
Sbjct 121 GTCATGGCCGCCCATCCCGGAGCTCATGCAGGAGGACGTCAGATCGTTCCTCTG 177
    
```

(c)

Fig. 4. BLASTN to identify mutation in coding region of *Waxy* gene

(a): Mutation(s) in exon 3; (b): Mutation(s) in exon 4; (c): Mutation(s) in exon 9

(Note: Query- mutant type; Subject- original type)

IDENTIFICATION OF SOME NUCLEOTIDE MUTATIONS IN WAXY GENE (BGIOSGA022241)...

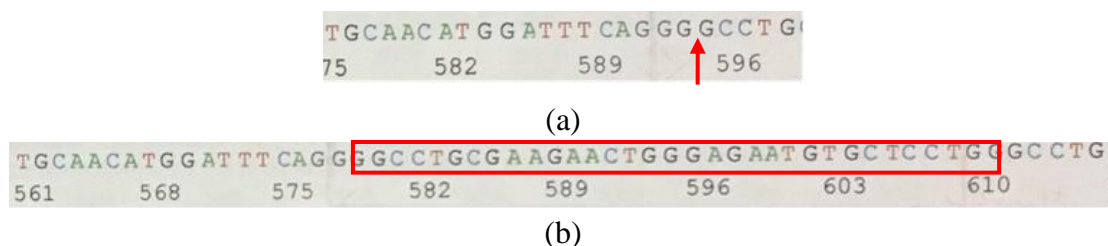


Fig. 5. The insertion of 32 nucleotides at the splicing point of intron 12
(a): original sequences; (b): mutant sequences

Total 3480 nucleotides of *Waxy* gene were analyzed via BLASTN and the result was shown in table II.

Based on these point mutations, new DNA marker was developed to improve efficiency of rice mutation breeding (Table III).

D. Development of new DNA marker for rice mutation breeding

Table II. The discovery of mutation in *Waxy* gene through BLASTN.

Gene region	Total	Identities (%)	Gaps (%)	Reference
Exon	1810	1806 (99,8%)	4 (0,2%)	- Exon 3: 34 (T/-); 71 (-/T) - Exon 4: 14 (C/T) - Exon 9: 115 (T/C)
Intron	1670	1611 (96,5%)	59 (3,5%)	- Intron 3: 29 (T/-); 31 (T/-) - Intron 5: 9 (T/C) - Intron 6: Exon6/intron6 junction (G/-); 53 (T/-); 59 (T/-); 63 (T/-) - Intron 8: 29 (A/G); 46 (-/T) - Intron 9: 81 (A/G); 95 (A/G); 99 (-/TAA); 139 (G/A); 142 (A/G); 148 (C/T); 161 (A/G); 165 (C/T); 177 (G/C); 193 (G/A) - Intron 11: 41 (T/C); 58 (A/G) - Intron 12: 83 (A/T); 98 (G/A); 134 (A/C); intron12/exon13 junction (insertion of 32 nucleotides)
Total	3480	3417 (98,2%)	63 (1,8%)	

Table III. The information of new developed DNA marker.

Sequence (5'- 3')	Target mutation	Annealing temperature	Expected size
W _x -F: GATTT C AGGTTTGGGGAAAGAT T	Nucleotide T at position 14 in exon 4	49.4 °C	1271 bp
W _x -R: TGCGGCGGCCATGACGTCAG G	Nucleotide C at position 115 in exon 9		

(Bold and underline character – mutation point)

IV. DISSCUSION

A. Amplify and sequence the full length of *Waxy* gene

Sixteen primers were designed in Table I with lengths from 24 to 25 nucleotides. The Wx-1F primer was designed at boundary of 5'-UTR/exon 1 and the Wx-8R primer was designed at boundary of exon 13/3'-UTR. There was no failure in amplifying *Waxy* genes of both original variety and its mutant by Wx-1F and Wx-8R. It was indicated that there was no difference at junction sites. In agarose gel, there is only one band of PCR products and this band is bold and densitic (Fig. 2). These criteria are very important for the accuracy of sequencing.

Results in Fig. 3 were good at reading: no sequences were miss-calls (N), high concentration, no spaced peaks, only one color for each peak and lack of baseline (noise). The full *Waxy* genes of original type and mutant type were sequenced successfully by sixteen primers (Table I).

B. Identify mutation in *Waxy* gene between the original type and its mutant type

Sequences of thirteen exons (coding regions) and twelve introns (non-coding regions) of *Waxy* gene from original and mutant lines were compared via BLASTN to identify mutation(s). The result in table II shown that, mutant rate in the non-coding region (3,5%) was higer than that in the coding region (0,2%). In coding region, there were 1806 identities (99.8%) and 4 gaps (0.2%) in coding region. Four gaps mean point mutations including: the deletion of T nucleotide (T/-) at point 34 and insertion of T (-/T) between points 70 and 71 (in exon 3); the substitution (C/T) at position 14 in exon 4 and the substitution T/C at position 115 in exon 9. In

total of 1670 non-coding nucleotides compared, it was shown 1611 identities (96.5%) and 59 gaps (3.5%) (table II). The changes were listed: deletions (T/-) at positions 29 and 31 in intron 3; the change (T/C) at position 9 in intron 5; the deletions (T/-) at position 53, 59 and the deletion (A/-) at the position 63 of intron 6; the change (A/G) at position 29 and the insertion (-/T) between positions 45 and 46 in intron 8; the substitutions (A/G) at positions 81, 95, 142, 161, the changes (G/A) at positions 139 and 193, the changes (C/T) at positions 148 and 165, the change (G/C) at position 177 and the insertion (-/TAA) between positions 98 and 99 in intron 9; the alterations (T/C) at position 41 and (A/G) at position 58 in intron 11; the alterations (A/T) at position 83, (G/A) at position 98, (A/C) at position 134, (G/T) at position 206 and the addition of 32 nucleotides "GGGCCTGCGAAGAAGACTGGGAGAATGT GCTCCT" at the end of intron 12.

Four point mutations collected in coding regions (exons) (Fig. 4) will result the effect on translation directly. Because information of proteins for life is coded by triplets, thus with every mRNA there are three frame of translation. In theoretical, the structure of DNA is double strands, thus there are total six frame of reading. Based on the C/T mutation at position 14 in exon 4 will cause the replacement of "T" in the original type to "I" the in mutant type; or "P" to "S". The T/C mutation at position 115 in exon 9 resulted substitution of amino acid sequences "XAXNKX" in original type to "KALNKE" in mutant type; or "XXXTRX" to "RR*TRR"; or "XXX" to "GAE". Mutations in exon 3, the deletion of T nucleotide at point 34 and insertion of T at point between 70 and 71, will create the change of amino acids starting from the mutant site.

In total 59 gaps identified in non-coding regions, two types of mutation with more frequency than other ones were the deletion (T/-) (with five observations) and the substitution (A/G) (with six observations). Mutations at intron/exon junctions were also determined and listed: the deletion G/- at the first of intron 6 and the insertion of 32 nucleotides at the end of intron12 (fig. 5). These results leading to us the next research to interpret that if these changes are effective on the cutting of intron 6 and intron 12 or not and how they regulate on the amylose content.

C. Development of new DNA marker for rice breeding relevant to amylose content

The forward primer Wx-F was designed based on the substitution C/T at position 14 in exon 4 with the length of 22 nucleotides and 40.9% GC content. The reverse primer Wx-R was designed based on the substitution T/C at position 115 in exon 9 with the length of 22 nucleotides and 72.7% GC content. The expected size of PCR product which is amplified by this new primer pair is 1271 bp and the recommended annealing temperature is 49.4°C (table III).

The new developed DNA marker which was designed with both point mutations at 3' of two primers in pair: the forward primer Wx-F (5'- GATTCAGGTTTGGGGAAAGAT**T** - 3') with the change C/T at position 14 in exon4 (nucleotide T – bold and underline) and the reverse primer Wx-R (5' - TGGCGGCGCCATGACGTCAG**G** - 3') with the substitution T/C at position 115 in exon 9 (nucleotide G – bold and underline). The 3' of primer which will be bind to the DNA strand firstly in transcription is better in conservating. Thus, the mutations were set in the first triplet of 3' to engage of the accuracy of mutant screening in PCR. This new developed DNA

marker will be studied in further by being used back directly for its mutant population before applying for selection.

V. CONCLUSIONS

(1) Four point mutations in coding regions (exon 3, exon 4 and exon 9) of *Waxy* gene would lead to the difference of amino acids in polypeptide in obvious.

(2) Some alterations at the first of intron 6 and the end of intron 12 will be done in more experiments to clarify their impact on expression of *Waxy* gene.

(3) It is important to study, utilize these mutants and new developed DNA marker to improve the efficiency of rice breeding with low amylose content.

ACKNOWLEDGMENT

This experiment was done at the Wakasa-wan Energy Research Center, Fukui, Japan, with the support of the Fukui International Human Resources Development Center for Atomic Energy (FIHRDC) FY 2016.

REFERENCES

- [1] S. X. Tang, G.S. Khush, and B.O. Juliano, "Variation and correlation of four cooking and eating quality indices of rices". Philipp Journal Crop Science, 14, 45-49, 1989.
- [2] P. D. Larkin and W. D. Park, "Association of *Waxy* gene single nucleotit polymorphisms with starch characteristics in rice (*Oryza sativa* L.)". Molecular Breeding, 12 (4), 335–339, 2003.
- [3] M. Nakagahara and T. Nagamine, "Spontaneous occurrence of low amylose genes and geographical distribution of amylose content in Asian rice". Rice Genetics Newsletter, 3, 46-48, 1986.

- [4] L. Liu, X. Ma, S. Liu, C. Zhu, L. Jiang, Y. Wang, Y. Shen, Y. Ren, H. Dong, L. Chen, X. Liu, Z. Zhao, H. Zhai, J. Wan, "Identification and characterization of a novel Waxy allele from Yunnan rice landrace". *Plant Molecular Biology*, 71, 609–626, 2009.
- [5] Kharabian Ardashir Masouleh, Daniel L. E. Waters, Russell F. Reinke, Rachele Ward & Robert J. Henry, "SNP in starch biosynthesis genes associated with nutritional and functional properties of rice". *Scientific Reports*, 2, Article number: 557, 2012.
- [6] M. H. Chen, C. J. Bergman, S. R. M. Pinson, R. G. Fjellstrom, "Waxy gene haplotypes: Associations with apparent amylose content and the effect by the environment in an international rice germplasm collection". *Journal of Cereal Science*, 47(3), 536-545, 2008.
- [7] Cheng Zai Quan, Liu Yan Ping, Chen Rui, Peng Bo, Xiong Hua Bin, Zhang Cheng, Zhong Qiao Fang and Huang Xing Qi, "Diversity of Waxy gene alleles in the wild rice species of the *Oryza* genus". *Botanical Studies*, 51, 403-411, 2010.
- [8] H. Y. Hirano, Y. Sano, "Molecular Characterization of the Waxy Locus of Rice (*Oryza sativa*)". *Plant and Cell Physiology*, 32 (7), 989-997, 1991.
- [9] I. Mikami, N. Uwatoko, Y. Ikeda, J. Yamaguchi, H. Y. Hirano, Y. Suzuki and Y. Sano, "Allelic diversification at the Wx locus in landraces of Asian rice". *Theoretical and Applied Genetics*, 116 (7), 979–89, 2008
- [10] M. Isshiki, K. Morino, M. Nakajima, R. J. Okagaki, S. R. Wessler, T. Izawa and K. Shimamoto, "A naturally occurring functional allele of the rice Waxy locus has a GT to TT mutation at the 5' splice site of the first intron". *The Plant Journal*, 15 (1), 133–138, 1998.
- [11] Y. Sano, "Differential regulation of Waxy gene expression in rice endosperm". *Theoretical and Applied Genetics*, 68 (5), 467-473, 1985.
- [12] X. L. Cai, Z. Y. Wang, Y. Y. Xing, J. L. Zhang, M. M. Hong, "Aberrant splicing of intron 1 leads to the heterogeneous 5' UTR and decreased expression of Waxy gene in rice cultivars of intermediate amylose content". *The Plant Journal*, 14(4), 459-465, 1998.
- [13] Z. Y. Wang, F. Q. Zheng, G. Z. Shen, J. P. Gao, D. P. Snustad, M. G. Li, J. L. Zhang, M. M. Hong, "The amylose content in rice endosperm is related to the post-transcriptional regulation of the Waxy gene". *The Plant Journal*, 7(4), 613-622, 1995.
- [14] M. Dobo, N. Ayres, G. Walker, W. D. Park, "Polymorphism in the GBSS gene affects amylose content in US and European rice germplasm". *Journal Cereal Science*, 52(3), 450–456, 2010.
- [15] N. M. Ayres, A. M. Mc Clung, P. D. Larkin, H. F. J. Bligh, C. A. Jones, W. D. Park, "Microsatellites and a single-nucleotide polymorphism differentiate apparent amylose classes in an extended pedigree of US rice germplasm". *Theoretical and Applied Genetics*, 94, 773–781, 1997.
- [16] C. Biselli, D. Cavalluzzo, R. Perrini, A. Gianinetti, P. Bagnaresi, S. Urso, G. Orasen, F. Desiderio, E. Lupotto, L. Cattivelli, "Improvement of marker-based predictability of Apparent Amylose Content in japonica rice through GBSSI allele mining". *Rice*, 7 (1), 2014.
- [17] P. D. Larkin and W. D. Park, "Transcript accumulation and utilization of alternate and non-consensus splice sites in rice granule-bound starch synthase are temperature-sensitive and controlled by a single-nucleotide polymorphism". *Plant Molecular Biology*, 40 (4), 719–727, 1999.
- [18] Tran Thi Thu Hoai, Hiroaki Matsusaka, Yoshiko Toyosawa, Tran Danh Suu, Hikaru Satoh and Toshihiro Kumamaru, "Influence of single-nucleotide polymorphisms in the gene encoding granule-bound starch synthase I on amylose content in Vietnamese rice cultivars". *Breeding science*, 64(2), 142–148, 2014.

IDENTIFICATION OF SOME NUCLEOTIDE MUTATIONS IN WAXY GENE (BGIOGA022241)...

- [19] A. Kharabian, “An efficient computational method for screening functional SNPs in plants”. *Journal of Theoretical Biology*, 265, 55–6, 2010.
- [20] H. Sato, Y. Suzuki, M. Sakai, T. Imbe, “Molecular characterization of Wx-mq, a novel mutant gene for low-amylose content in endosperm of rice (*Oryza sativa* L.)”. *Breeding Science*, 52, 131–135, 2002.
- [21] J. S. Bao, H. Corke, M. Sun, “Nucleotide diversity in starch synthase IIa and validation of single nucleotide polymorphisms in relation to starch gelatinization temperature and other physicochemical properties in rice (*Oryza sativa* L.)”. *Theoretical and Applied Genetics*, 113, 1171–1183, 2006.
- [22] T. T. Hoai, A. Nishi and H. Satoh, “Diversity of granule bound starch synthesis (GBSS) levels in North Vietnam local rice cultivars”. *Rice Genetics Newsletter*, 24, 62–64, 2008.
- [23] M. S. Jahan, T. Kumamaru, A. Hamid and H. Satoh, “Diversity of granule bound starch synthase (GBSS) level in Bangladesh rice cultivars”. *Rice Genetics Newsletter*, 19, 69–71, 2002.
- [24] H. Satoh, R. X. Ronald and T. C. Katayama, “On amylose content of cultivated rice collected in Madagasca, Kagoshima University Research Center South Pacific”, *Occasional Papers*, 18, 83–91, 1990
- [25] <https://www.qiagen.com>
- [26] <http://www.gramene.org>

APPENDIX

The comparison between *Waxy* genes of original and mutant types

Exon1

Score	Expect	Identities	Gaps	Strand
627 bits(339)	0.0	339/339(100%)	0/339(0%)	Plus/Plus
Query 1	ATGTCGGCTCTCACCACGTCCCAGCTCGCCACCTCGGCCACCGGCTTCGGCATCGCCGAC	60		
Sbjct 1	ATGTCGGCTCTCACCACGTCCCAGCTCGCCACCTCGGCCACCGGCTTCGGCATCGCCGAC	60		
Query 61	AGGTCGGCGCCGTCGTCGCTGCTCCGCCACGGGTTCAGGGCCTCAAGCCCCGAGCCCC	120		
Sbjct 61	AGGTCGGCGCCGTCGTCGCTGCTCCGCCACGGGTTCAGGGCCTCAAGCCCCGAGCCCC	120		
Query 121	GCCGGCGGCGACGCGACGTCGCTCAGCGTGACGACCAGCGCGCGCGACGCCAAGCAG	180		
Sbjct 121	GCCGGCGGCGACGCGACGTCGCTCAGCGTGACGACCAGCGCGCGCGACGCCAAGCAG	180		
Query 181	CAGCGGTCGGTGCAGCGTGGCAGCCGGAGGTTCCCTCCGTCGTCGTGTACGCCACCGGC	240		
Sbjct 181	CAGCGGTCGGTGCAGCGTGGCAGCCGGAGGTTCCCTCCGTCGTCGTGTACGCCACCGGC	240		
Query 241	GCCGGCATGAACGTCGTGTTTCGTCGGCGCCGAGATGGCCCCCTGGAGCAAGACCGGCGGC	300		
Sbjct 241	GCCGGCATGAACGTCGTGTTTCGTCGGCGCCGAGATGGCCCCCTGGAGCAAGACCGGCGGC	300		
Query 301	CTCGGTGACGTCCTCGGTGGCCTCCCCCTGCCATGGCT	339		

Sbjct 301 |||||
CTCGGTGACGTCTCTCGGTGGCCTCCCCCTGCCATGGCT 339

Exon 2

Score	Expect	Identities	Gaps	Strand
150 bits(81)	2e-42	81/81(100%)	0/81(0%)	Plus/Plus
Query 1	GCGAATGGCCACAGGGTCATGGT	GATCTCTCCTCGGTACGACCAGTACAAGGACGCTTGG		60
Sbjct 1	GCGAATGGCCACAGGGTCATGGT	GATCTCTCCTCGGTACGACCAGTACAAGGACGCTTGG		60
Query 61	GATACCAGCGTTGTGGCTGAG		81	
Sbjct 61	GATACCAGCGTTGTGGCTGAG		81	

Exon 3

Score	Expect	Identities	Gaps	Strand
174 bits(94)	2e-49	99/101(98%)	2/101(1%)	Plus/Plus
Query 1	ATCAAGGTTGCAGACAGGTACGAGAGGGTGAGG	-TTTTTCCATTGCTACAAGCGTGGAGT		59
Sbjct 1	ATCAAGGTTGCAGACAGGTACGAGAGGGTGAGG	TTTTTCCATTGCTACAAGCGTGGAGT		60
Query 60	CGACCGTGTG	TTCATCGACCATCCGTCATT	CCTGGAGAAG	100
Sbjct 61	CGACCGTGTG	TTCATCGACCATCCGTCATT	CCTGGAGAAG	100

Exon 4

Score	Expect	Identities	Gaps	Strand
161 bits(87)	1e-45	89/90(99%)	0/90(0%)	Plus/Plus
Query 1	GTTTGGGGAAAGA	TTCGGAGAGAAGATCTACGGACCTGACACTGGAGTTGATTACAAAGAC		60
Sbjct 1	GTTTGGGGAAAGA	CTCGGAGAGAAGATCTACGGACCTGACACTGGAGTTGATTACAAAGAC		60
Query 61	AACCAGATGCGTTTCAGCCTTCTTTGCCAG		90	
Sbjct 61	AACCAGATGCGTTTCAGCCTTCTTTGCCAG		90	

Exon 5

Score	Expect	Identities	Gaps	Strand
119 bits(64)	4e-33	64/64(100%)	0/64(0%)	Plus/Plus
Query 1	GCAGCACTCGAGGCTCCTAGGATCCTAAACCTCAACAACAACCCATACTTCAAAGGAACT			60
Sbjct 1	GCAGCACTCGAGGCTCCTAGGATCCTAAACCTCAACAACAACCCATACTTCAAAGGAACT			60

IDENTIFICATION OF SOME NUCLEOTIDE MUTATIONS IN WAXY GENE (BGIOSGA022241)...

```
Query 61 TATG 64
      ||||
Sbjct 61 TATG 64
```

Exon 6

Score	Expect	Identities	Gaps	Strand
187 bits(101)	3e-53	101/101(100%)	0/101(0%)	Plus/Plus

```
Query 1 GTGAGGATGTTGTGTTTCGTCTGCAACGACTGGCACACTGGCCCACTGGCGAGCTACCTGA 60
      ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Sbjct 1 GTGAGGATGTTGTGTTTCGTCTGCAACGACTGGCACACTGGCCCACTGGCGAGCTACCTGA 60
```

```
Query 61 AGAACAACTACCAGCCCAATGGCATCTACAGGAATGCAAAG 101
      ||||||||||||||||||||||||||||||||||||||
Sbjct 61 AGAACAACTACCAGCCCAATGGCATCTACAGGAATGCAAAG 101
```

Exon 7

Score	Expect	Identities	Gaps	Strand
226 bits(122)	9e-65	122/122(100%)	0/122(0%)	Plus/Plus

```
Query 1 TTTTCACTGCAGGTTGCTTTCTGCATCCACAACATCTCCTACCAGGGCCGTTTCGCTTTC 60
      ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Sbjct 1 TTTTCACTGCAGGTTGCTTTCTGCATCCACAACATCTCCTACCAGGGCCGTTTCGCTTTC 60
```

```
Query 61 GAGGATTACCCTGAGCTGAACCTCTCCGAGAGGTTCAGGTCATCCTTCGATTTTCATCGAC 120
      ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Sbjct 61 GAGGATTACCCTGAGCTGAACCTCTCCGAGAGGTTCAGGTCATCCTTCGATTTTCATCGAC 120
```

```
Query 121 GG 122
      ||
Sbjct 121 GG 122
```

Exon 8

Score	Expect	Identities	Gaps	Strand
451 bits(244)	6e-132	244/244(100%)	0/244(0%)	Plus/Plus

```
Query 1 GTATGACACGCCGGTGGAGGGCAGGAAGATCAACTGGATGAAGGCCGGAATCCTGGAAGC 60
      ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Sbjct 1 GTATGACACGCCGGTGGAGGGCAGGAAGATCAACTGGATGAAGGCCGGAATCCTGGAAGC 60
```

```
Query 61 CGACAGGGTGCTCACCCTGAGCCCGTACTACGCCGAGGAGCTCATCTCCGGCATCGCCAG 120
      ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Sbjct 61 CGACAGGGTGCTCACCCTGAGCCCGTACTACGCCGAGGAGCTCATCTCCGGCATCGCCAG 120
```

```
Query 121 GGGATGCGAGCTCGACAACATCATGCGGCTCACCGGCATCACCGGCATCGTCAACGGCAT 180
```

```

|||||
Sbjct 121 GGGATGCGAGCTCGACAACATCATGCGGCTCACCGGCATCACCGGCATCGTCAACGGCAT 180

Query 181 GGACGTCAGCGAGTGGGATCCCAGCAAGGACAAGTACATCACCGCCAAGTACGACGCAAC 240
|||||
Sbjct 181 GGACGTCAGCGAGTGGGATCCCAGCAAGGACAAGTACATCACCGCCAAGTACGACGCAAC 240

Query 241 CACG 244
||||
Sbjct 241 CACG 244

```

Exon 9

Score	Expect	Identities	Gaps	Strand
322 bits(174)	2e-93	176/177(99%)	0/177(0%)	Plus/Plus
Query 1	GCAATCGAGGCGAAGGCGCTGAACAAGGAGGCGTTGCAGGCGGAGGCGGGTCTTCCGGTC			60
Sbjct 1	GCAATCGAGGCGAAGGCGCTGAACAAGGAGGCGTTGCAGGCGGAGGCGGGTCTTCCGGTC			60
Query 61	GACAGGAAAATCCCACTGATCGCGTTCATCGGCAGGCTGGAGGAACAGAAGGGC <u>C</u> CTGAC			120
Sbjct 61	GACAGGAAAATCCCACTGATCGCGTTCATCGGCAGGCTGGAGGAACAGAAGGGC <u>T</u> CTGAC			120
Query 121	GTCATGGCCCGCCATCCCGAGCTCATGCAGGAGGACGTCCAGATCGTTCTTCTG			177
Sbjct 121	GTCATGGCCCGCCATCCCGAGCTCATGCAGGAGGACGTCCAGATCGTTCTTCTG			177

Exon 10

Score	Expect	Identities	Gaps	Strand
355 bits(192)	3e-103	192/192(100%)	0/192(0%)	Plus/Plus
Query 1	GGTACTGGAAAGAAGAAGTTCGAGAAGCTGCTCAAGAGCATGGAGGAGAAGTATCCGGGC			60
Sbjct 1	GGTACTGGAAAGAAGAAGTTCGAGAAGCTGCTCAAGAGCATGGAGGAGAAGTATCCGGGC			60
Query 61	AAGGTGAGGGCCGTGGTGAAGTTCAACGCGCCGCTTGCTCATCTCATCATGGCCGGAGCC			120
Sbjct 61	AAGGTGAGGGCCGTGGTGAAGTTCAACGCGCCGCTTGCTCATCTCATCATGGCCGGAGCC			120
Query 121	GACGTGCTCGCCGTCCCCAGCCGCTTCGAGCCCTGTGGACTCATCCAGCTGCAGGGGATG			180
Sbjct 121	GACGTGCTCGCCGTCCCCAGCCGCTTCGAGCCCTGTGGACTCATCCAGCTGCAGGGGATG			180
Query 181	AGATACGGAACG 192			
Sbjct 181	AGATACGGAACG 192			

IDENTIFICATION OF SOME NUCLEOTIDE MUTATIONS IN WAXY GENE (BGIOGA022241)...

Exon 11

Score	Expect	Identities	Gaps	Strand
161 bits(87)	1e-45	87/87(100%)	0/87(0%)	Plus/Plus
Query 1	CCCTGTGCTTGC	CGTCCACCGGTGGGCTCGTGGACACGGTCATCGAAGGCAAGACTGGT	60	
Sbjct 1	CCCTGTGCTTGC	CGTCCACCGGTGGGCTCGTGGACACGGTCATCGAAGGCAAGACTGGT	60	
Query 61	TTCCACATGGG	CCGCTCTCAGCGTCGAC 87		
Sbjct 61	TTCCACATGGG	CCGCTCTCAGCGTCGAC 87		

Exon 12

Score	Expect	Identities	Gaps	Strand
239 bits(129)	1e-68	129/129(100%)	0/129(0%)	Plus/Plus
Query 1	TGCAAGGTGGT	TGGAGCCAAGCGACGTGAAGAAGGTGGCGGCCACCCTGAAGCGCGCCATC	60	
Sbjct 1	TGCAAGGTGGT	TGGAGCCAAGCGACGTGAAGAAGGTGGCGGCCACCCTGAAGCGCGCCATC	60	
Query 61	AAGGTCGTC	CGGCACGCCGGCGTACGAGGAGATGGTCAGGAACTGCATGAACCAGGACCTC	120	
Sbjct 61	AAGGTCGTC	CGGCACGCCGGCGTACGAGGAGATGGTCAGGAACTGCATGAACCAGGACCTC	120	
Query 121	TCCTGGAAG	129		
Sbjct 121	TCCTGGAAG	129		

Exon 13

Score	Expect	Identities	Gaps	Strand
158 bits(85)	2e-44	85/85(100%)	0/85(0%)	Plus/Plus
Query 1	GGGCCTGGG	CGTCGCCGGCAGCGCGCCGGGATCGAAGGCGACGAGATCGCGCCGCTCGC	60	
Sbjct 1	GGGCCTGGG	CGTCGCCGGCAGCGCGCCGGGATCGAAGGCGACGAGATCGCGCCGCTCGC	60	
Query 61	CAAGGAGA	ACGTGGCTGCTCCTTGA 85		
Sbjct 61	CAAGGAGA	ACGTGGCTGCTCCTTGA 85		

Intron 1

Score	Expect	Identities	Gaps	Strand
209 bits(113)	8e-60	113/113(100%)	0/113(0%)	Plus/Plus
Query 1	GTAAGCACAC	ACAACTTCGATCGCTCGTCGTCGCTGACCGTCGTCGCTTCAACTGTTC	60	
Sbjct 1	GTAAGCACAC	ACAACTTCGATCGCTCGTCGTCGCTGACCGTCGTCGCTTCAACTGTTC	60	

Query 61 TTGATCATCGCATTGGATGGATGTGTAATGTTGTGTTCTTGTGTTCTTTGCAG 113
 |||
 Sbjct 61 TTGATCATCGCATTGGATGGATGTGTAATGTTGTGTTCTTGTGTTCTTTGCAG 113

Intron 2

Score	Expect	Identities	Gaps	Strand
198 bits(107)	2e-56	107/107(100%)	0/107(0%)	Plus/Plus

Query 1 GTAGGAGCATATGCGTGATCAGATCATCACAAGATCGATTAGCTTTAGATGATTTGTTAC 60
 |||
 Sbjct 1 GTAGGAGCATATGCGTGATCAGATCATCACAAGATCGATTAGCTTTAGATGATTTGTTAC 60

Query 61 ATTTGCAAGATTTTAACCCAAGTTTTGTGGTCAATTCATTGCAG 107
 |||
 Sbjct 61 ATTTGCAAGATTTTAACCCAAGTTTTGTGGTCAATTCATTGCAG 107

Intron 3

Score	Expect	Identities	Gaps	Strand
169 bits(91)	1e-47	96/98(98%)	2/98(2%)	Plus/Plus

Query 1 GTGGAGTCATCATTAGTTACCTttttt-g-tttttACTGAATTATTAACAGTGCATTTA 58
 ||| |
 Sbjct 1 GTGGAGTCATCATTAGTTACCTTTTTTTGTTTTTTACTGAATTATTAACAGTGCATTTA 60

Query 59 GCAGTTGGACTGAGCTTAGCTTCCACTGGTGATTTTCAG 96
 |||
 Sbjct 61 GCAGTTGGACTGAGCTTAGCTTCCACTGGTGATTTTCAG 98

Intron 4

Score	Expect	Identities	Gaps	Strand
182 bits(98)	1e-51	98/98(100%)	0/98(0%)	Plus/Plus

Query 1 GTCAGTGATTACTTCTATCTGATGATGGTTGGAAGCATCACGAGTTTACCATAGTATGTA 60
 |||
 Sbjct 1 GTCAGTGATTACTTCTATCTGATGATGGTTGGAAGCATCACGAGTTTACCATAGTATGTA 60

Query 61 TGGATTCACTAATAATTCGTGTATTGATGCTACTGCAG 98
 |||
 Sbjct 61 TGGATTCACTAATAATTCGTGTATTGATGCTACTGCAG 98

Intron 5

Score	Expect	Identities	Gaps	Strand
165 bits(89)	1e-46	91/92(99%)	0/92(0%)	Plus/Plus

Query 1 GTGAGTTACAAATTGATCTCAAGATCTTATAACTTTCTCGAAGGAATCCATGATGATCAG 60
 |||

IDENTIFICATION OF SOME NUCLEOTIDE MUTATIONS IN WAXY GENE (BGIOSGA022241)...

```

Sbjct 1 GTGAGTTATAATTGATCTCAAGATCTTATAACTTTCTCGAAGGAATCCATGATGATCAG 60

Query 61 ACTAATTCCTTCCGGTTTGTTACTGACAACAG 92
      |||||||||||||||||||||||||||||||
Sbjct 61 ACTAATTCCTTCCGGTTTGTTACTGACAACAG 92
  
```

Intron 6

Score	Expect	Identities	Gaps	Strand
130 bits(70)	3e-36	78/81(96%)	3/81(3%)	Plus/Plus

```

Query 1 GTCTATGCTTGTTCTTGCCATACCAACTCAAATCTGCATGCACACTGCATT-CTGTT-CA 58
      ||||||||||||||||||||||||||||||||||||||||||||| |||||
Sbjct 2 GTCTATGCTTGTTCTTGCCATACCAACTCAAATCTGCATGCACACTGCATTTCTGTTTCA 61

Query 59 G-AAACTGACTGTCTGAATCT 78
      | |||||||||||||||
Sbjct 62 G-AAACTGACTGTCTGAATCT 82
  
```

Intron 7

Score	Expect	Identities	Gaps	Strand
224 bits(121)	3e-64	121/121(100%)	0/121(0%)	Plus/Plus

```

Query 1 GTATGAGTAAGATTCTAAGAGTAACTTACTGTCAATTCGCCATATATCGATTCAATCCAA 60
      |||||||||||||||||||||||||||||||||||||||||||||||
Sbjct 1 GTATGAGTAAGATTCTAAGAGTAACTTACTGTCAATTCGCCATATATCGATTCAATCCAA 60

Query 61 GATCCTTTTGAGCTGACAACCCTGCACTACTGTCCATCGTTCAAATCCGGTTAAATTTCA 120
      |||||||||||||||||||||||||||||||||||||||||||||||
Sbjct 61 GATCCTTTTGAGCTGACAACCCTGCACTACTGTCCATCGTTCAAATCCGGTTAAATTTCA 120

Query 121 G 121
      |
Sbjct 121 G 121
  
```

Intron 8

Score	Expect	Identities	Gaps	Strand
204 bits(110)	4e-58	115/117(98%)	1/117(0%)	Plus/Plus

```

Query 1 GTAAGAACGAATGCATTCTTACAAGATGTGCAATCTGAATTTTCTTTGAAAAAGAAATT 60
      ||||||||||||||||||||||||| ||||||||||||| |||||||||||
Sbjct 1 GTAAGAACGAATGCATTCTTACAAGATATGCAATCTGAATTTTC-TTTGAAAAAGAAATT 59

Query 61 ATCATCTGTCACCTTCTTGATTGATTCTGACAAGGCAAGAATGAGTGACAAATTTTCAG 117
      |||||||||||||||||||||||||||||||||||||||||||||
  
```

Sbjct 60 ATCATCTGTCACTTCTTGATTGATTCTGACAAGGCAAGAATGAGTGACAAATTTTCAG 116

Intron 9

Score	Expect	Identities	Gaps	Strand
375 bits(203)	4e-109	231/243(95%)	3/243(1%)	Plus/Plus
Query 1	GTATAATATAATACACTACAAGACACACTTGCACGATATGCCAAAAATTCAGAACAAATT	60		
Sbjct 1	GTATAATATAATACACTACAAGACACACTTGCACGATATGCCAAAAATTCAGAACAAATT	60		
Query 61	CAGTGGCaaaaaaaaaCTC <u>G</u> AATATTAGGGAAG <u>G</u> ACC <u>TAA</u> TAATATCAAATAATTAGAA	120		
Sbjct 61	CAGTGGCAAAAAAAAAAACTC <u>A</u> AATATTAGGGAAG <u>A</u> ACC <u>---</u> TAATATCAAATAATTAGAA	117		
Query 121	GGGGTGAGGCTTTGAACCCAG <u>A</u> TC <u>G</u> TCTAG <u>T</u> CCACCACCTTGT <u>G</u> GAG <u>T</u> TAGCCGGAAGAG <u>C</u>	180		
Sbjct 118	GGGGTGAGGCTTTGAACCCAG <u>G</u> TC <u>A</u> TCTAG <u>C</u> CCACCACCTTGT <u>A</u> GAG <u>C</u> TAGCCGGAAGAG <u>G</u>	177		
Query 181	CTCTGAGCATTCTC <u>A</u> ATTCAGTGGCAAATGATGTGTATAATTTTGATCCGTGTGTGTTT	240		
Sbjct 178	CTCTGAGCATTCTC <u>G</u> ATTCAGTGGCAAATGATGTGTATAATTTTGATCCGTGTGTGTTT	237		
Query 241	CAG 243			
Sbjct 238	CAG 240			

Intron 10

Score	Expect	Identities	Gaps	Strand
196 bits(106)	5e-56	106/106(100%)	0/106(0%)	Plus/Plus
Query 1	GTATACAATTTCCATCTATCAATTCGATTGTTTCGATTTCATCTTTGTGCAATGCAATGCA	60		
Sbjct 1	GTATACAATTTCCATCTATCAATTCGATTGTTTCGATTTCATCTTTGTGCAATGCAATGCA	60		
Query 61	ATTGCAAATGCAAATGCATGATGATTTTCCTTGTTGATTTCTCCAG 106			
Sbjct 61	ATTGCAAATGCAAATGCATGATGATTTTCCTTGTTGATTTCTCCAG 106			

Intron 11

Score	Expect	Identities	Gaps	Strand
191 bits(103)	3e-54	107/109(98%)	0/109(0%)	Plus/Plus
Query 1	GTAAGCCTATACATTTACATAACAATCAGATATGACACAT <u>C</u> CTAATACCGATAAGT <u>C</u> GGT	60		

IDENTIFICATION OF SOME NUCLEOTIDE MUTATIONS IN WAXY GENE (BGIOGA022241)...

```
Sbjct 1   GTAAGCCTATACATTTACATAACAATCAGATATGACACATTCTAATACCGATAAGTCAGT 60
Query 61   ACACTACTACACATTTACATGGTTGCTGGTTATATGGtttttttGGCAG 109
          |||
Sbjct 61   ACACTACTACACATTTACATGGTTGCTGGTTATATGGTTTTTTGGCAG 109
```

Intron 12

Score	Expect	Identities	Gaps	Strand
638 bits(345)	0.0	353/357(99%)	0/357(0%)	Plus/Plus
Query 1	GTATAAATTACGAAACAAATTTAACCCAAACATATACTATATACTCCCTCCGCTTCTAAA	60		
Sbjct 1	GTATAAATTACGAAACAAATTTAACCCAAACATATACTATATACTCCCTCCGCTTCTAAA	60		
Query 61	TATTCAACGCCGTTGTCTTTTT <u>T</u> AAATATGTTTGACC <u>A</u> TTTCGTCTTATTaaaaaaTTAA	120		
Sbjct 61	TATTCAACGCCGTTGTCTTTTT <u>A</u> AAATATGTTTGACC <u>G</u> TTTCGTCTTATTAAAAAATTA	120		
Query 121	ATAATTATAAATT <u>C</u> TTTTCCATCATTGATTCAATTGTTAAATATACTTATATGTATACA	180		
Sbjct 121	ATAATTATAAATT <u>A</u> TTTTCCATCATTGATTCAATTGTTAAATATACTTATATGTATACA	180		
Query 181	TATAGTTTACATATTCATAAAA <u>T</u> TTTTGAACAAGACGAACGGTCAAACATGTGCTAA	240		
Sbjct 181	TATAGTTTACATATTCATAAAA <u>G</u> TTTTGAACAAGACGAACGGTCAAACATGTGCTAA	240		
Query 241	AAAGTTAACGGTGTGCAATATTCAGAAACGGAGGGAGTATAAACGTCTTGTTTCAGAAGTT	300		
Sbjct 241	AAAGTTAACGGTGTGCAATATTCAGAAACGGAGGGAGTATAAACGTCTTGTTTCAGAAGTT	300		
Query 301	CAGAGATTCACCTGTCTGATGCTGATGATGATTAATTGTTTGCAACATGGATTTTCAG	357		
Sbjct 301	CAGAGATTCACCTGTCTGATGCTGATGATGATTAATTGTTTGCAACATGGATTTTCAG	357		